

STUDIECENTRUM VOOR KERNENERGIE CENTRE D'ETUDE DE L'ENERGIE NUCLEAIRE

Geomicrobiology of waste repositories

Gas generation & removal Radionuclide migration & retention

Natalie Leys, Hugo Moors, Katinka Wouters,

Natalie.Leys@sckcen.be

Geomicrobiology in waste repositories

- Geochemical experimental set-ups were, are and will be influenced by microbiological activity.
 - E.g. Microbial H₂ consumption & CH₄ production in 'gas diffusion tests'
 - E.g. Microbial gas production in 'nitrate tests'
 - E.g. Biocorrosion and biodegradation of equipment & sensors
 - E.g. Microbial colonisation & biofilm development 'hampering in situ monitoring' and analysis of piezometers
 - ...

Repository and host rock were, are and will never be sterile.

(And for safety it is irrelevant whether the microbes are introduced or indigenous.)



Geochemistry

Geology & Geohydrology

Geomicrobiology

Functional and Safe waste disposal in repository

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Geomicrobiology in waste repositories

• Problem

Microbes will be present and active,

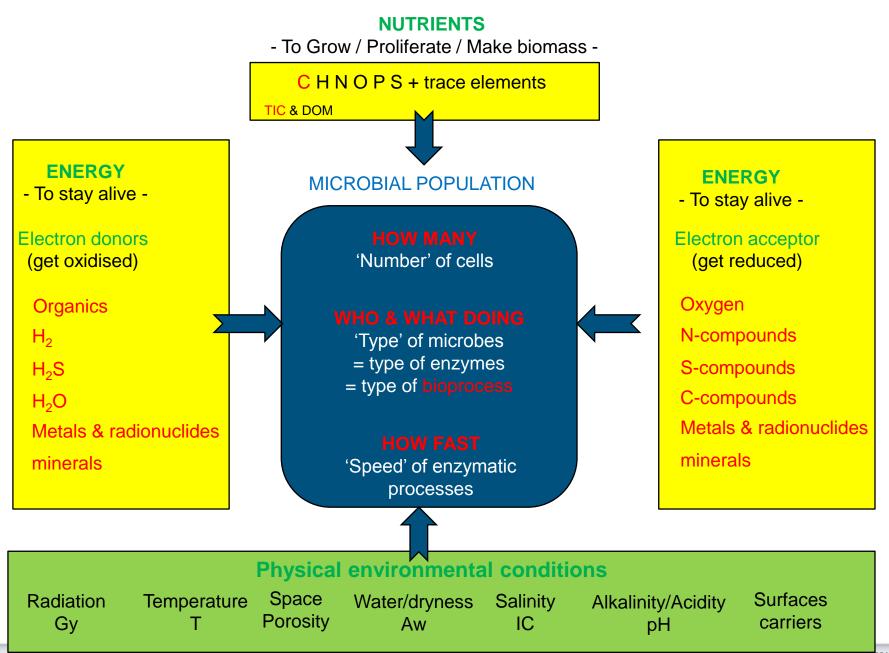
and will interact with waste, container, repository and host rock,

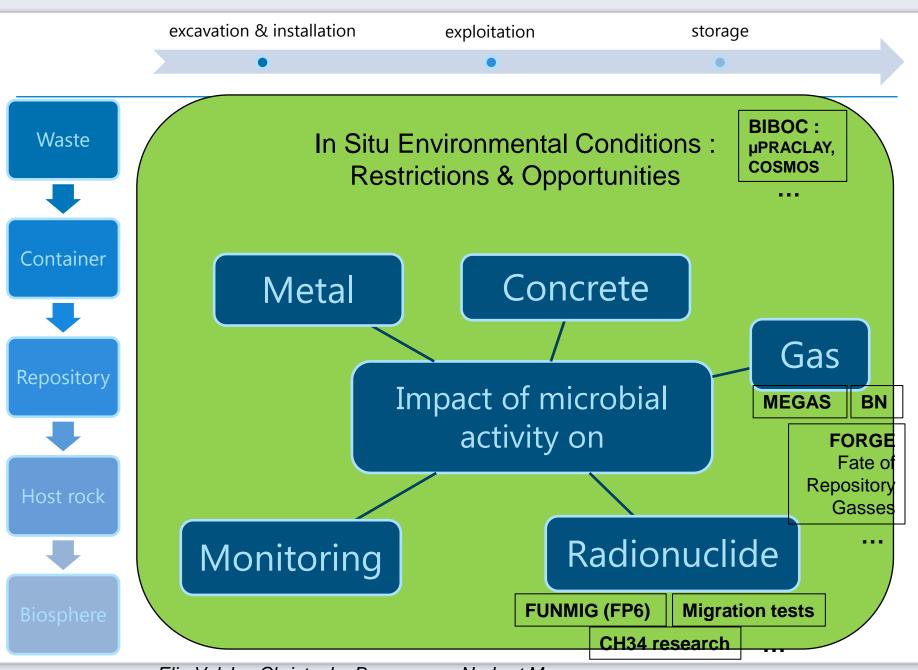
during excavation, exploitation, and storage.

Microbiology cannot be neglected but has to be considered and taken into account in the experimentation and modelling, and the final design and operational procedures, to assure functional and safe waste storage.

Need

- A better understanding of
 - the microbial populations present in waste, repository and host rock
 - the microbial bioprocesses that can occur at in situ conditions
 - the impact of those processes on the water & surface geochemistry
 - the impact of those geochemical changes on the waste, repository and host rock barrier function





Elie Valcke, Christophe Bruggeman, Norbert Maes,

Gas

Microbial Production of Gas

- Nitrogen cyclus :
 - nitrate from waste \rightarrow N₂O, NO, N₂

• Sulphur cyclus :

- clay \rightarrow pyrite \rightarrow sulphate (gips)
- sulphate (gips) \rightarrow H₂S

• Carbon cyclus :

- organic acids $\rightarrow CO_2$
- $CO_2 \rightarrow$ methane CH_4

(NRB, Nitrate <u>E-accep</u>tor, only when anoxic)

(during excavation, when oxic) (SRB, sulphate <u>E-accepto</u>r, only when anoxic)

(Hetrotrophs & organic E-donor, oxic & anoxic) (methanogens, CO_2 E-acceptor, anoxic)

Production of reduced gas species with impact on

- → water geochemistry
- → structural materials (corrosion)
- → dissolution chemistry and mobility of radionuclides

Gas

Microbial Consumption of Gas

Oxygen

- Most preferred E-acceptor for most microbes, can be consumed very fast
- Hydrogen from anoxic corrosion & radiolysis
 - very good 'fuel for microbes' (E-donor)
 - oxic (with O_2 as E-acceptor)
 - anoxic (with nitrate or sulphate as E-acceptor)

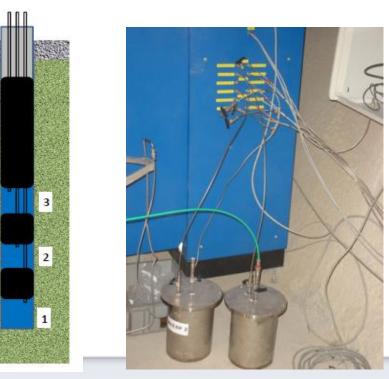
Carbon dioxide

As carbon source for autotrophic proliferation

e.g. BN-Experiment, Mont Terri

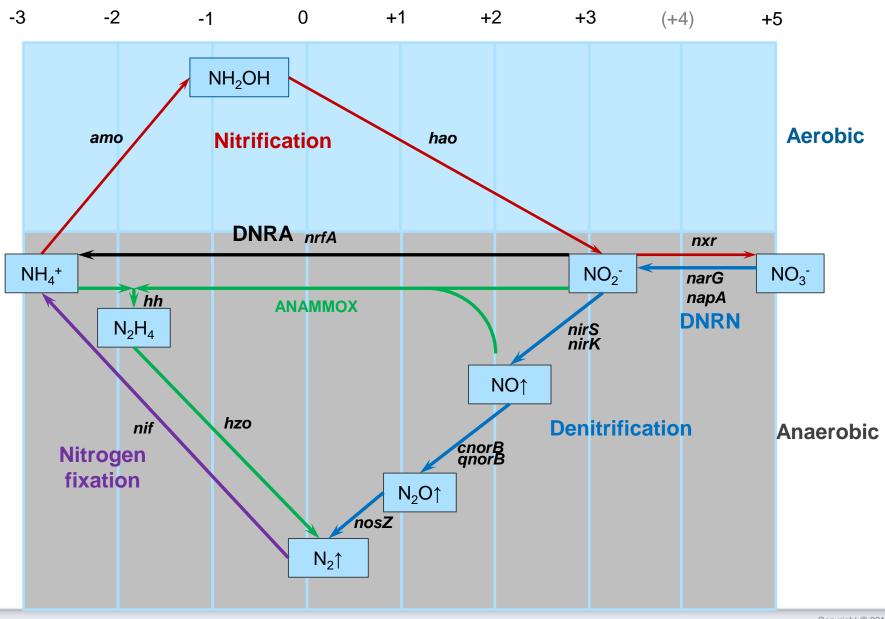
Interaction clay – bitumen waste Nitrate & Acetate injection as products

- 3 packed-off intervals
- Sintered stainless steel filter screen surrounding a central tube
- Water lines connecting intervals with recirculation cabinet in gallery
- \rightarrow Providing 'space' and 'water' & 'rock contact'

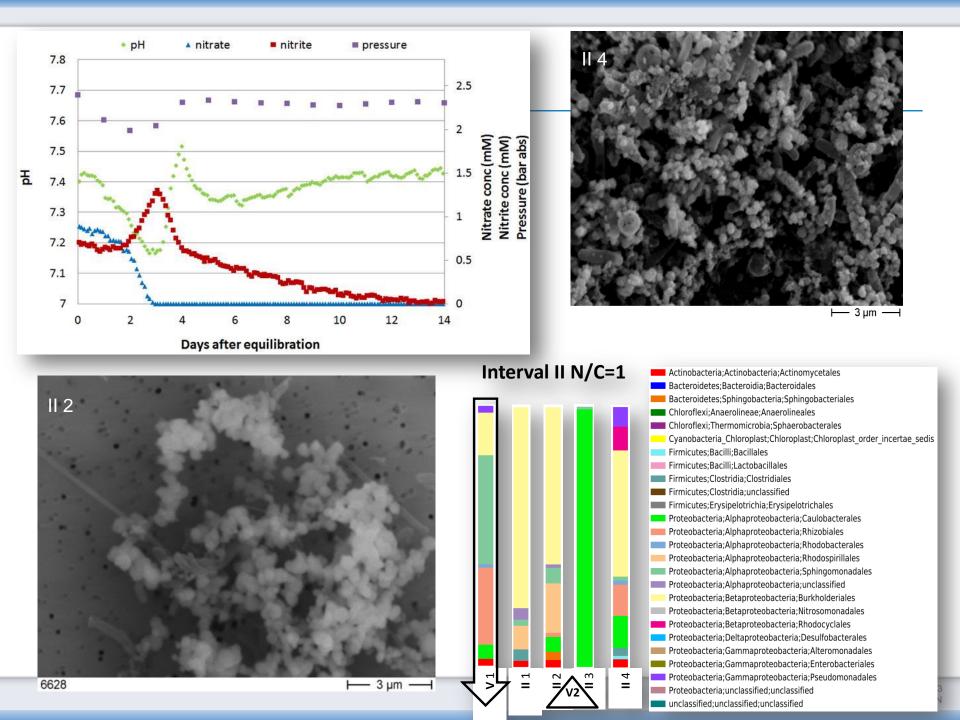




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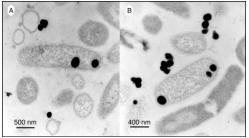
Radionuclide

Radionuclide migration

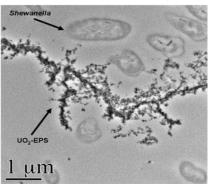
- RN : selenium (Se), uranium (U), neptunium (Np), plutonium (Pu), curium (Cm)
- Microbial impact on RN chemical form (speciation)
 - Microbial RN reduction (RN as E-acceptor, anoxic)
 → e.g. metalic nanoparticles
- Microbial impact on RN ligands
 - Degradation of organic ligands (as C-source)
 - Production of complexion compounds
 - biomineralisation with phosphate, carbonate, carboxylate ...
 - Extracellular polymeric substances (EPS)
 - metallophores (e.g. organic acids, peptides, ...)
- Microbial impact on RN sorption
 - RN sorption on microbial biofilm on wetted surfaces
 - RN sorption on microbial colloids

alkalinity (pH), redox potential (Eh),

Selenihalanaerobacter shriftii & selenate (A) and selenite (B) \rightarrow Black = solid elemental Se.



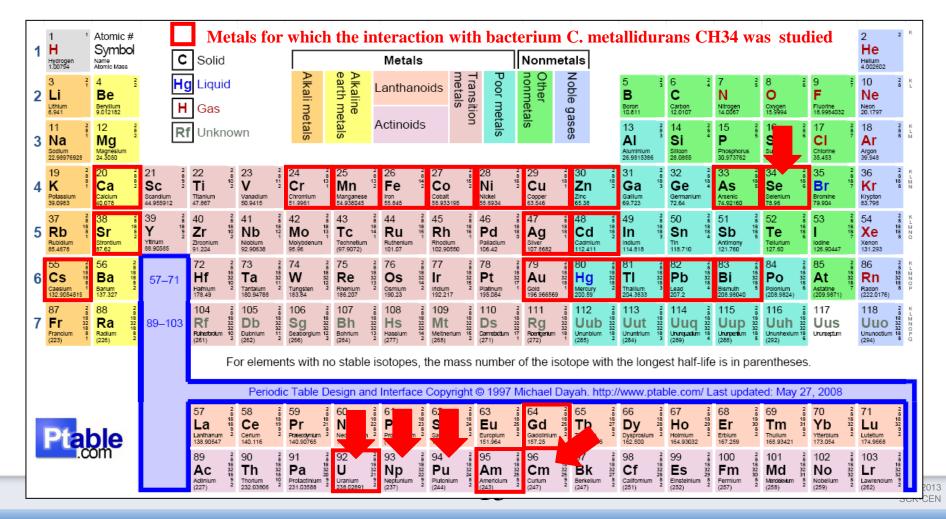
Shewanella & uranyl \rightarrow Black = solid uraninite (UO2).



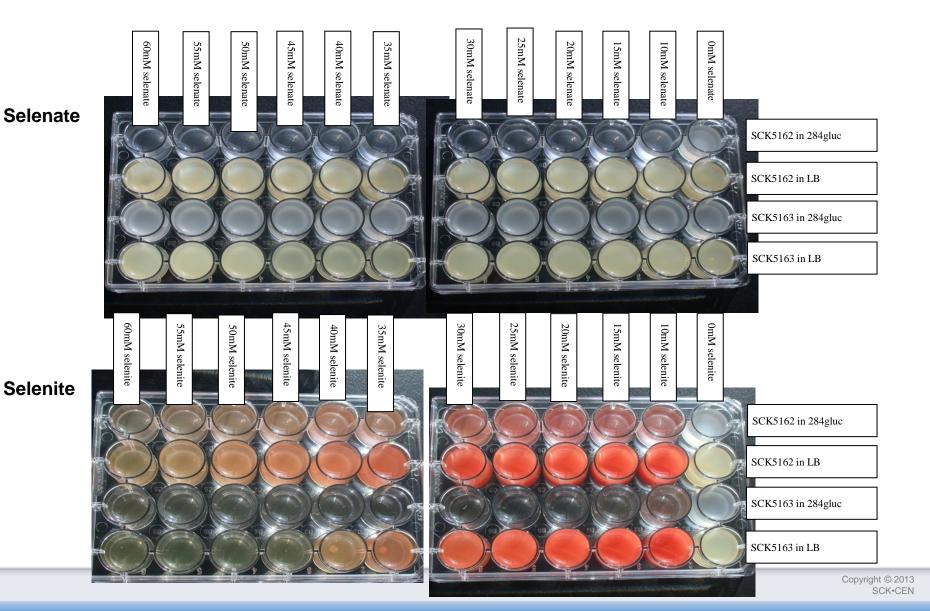
e.g. Characterising and 'engineering' bacteria that bioaccumulate or bioprecipitate metals & RN

Our model bacterium: Cupriavidus metallidurans CH34

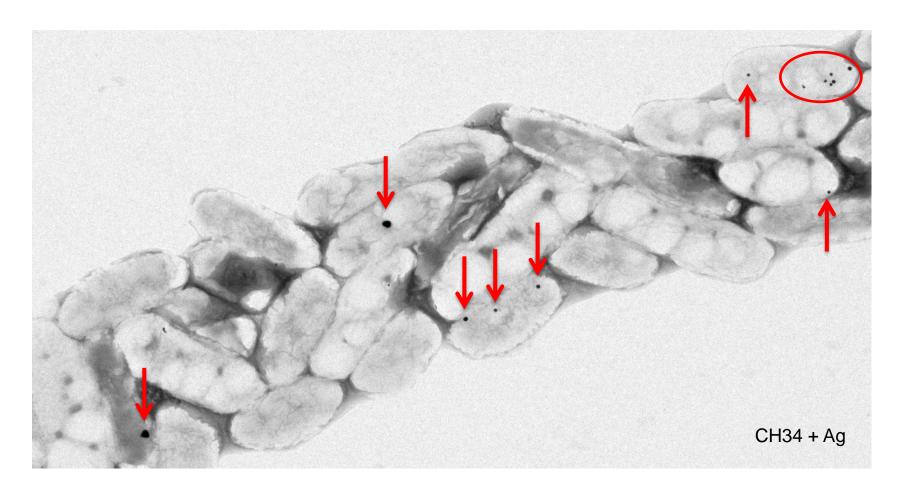
Isolated in 1976 from polluted sludge in metallurgical plant (Prayon-Liege, Belgium), by Dr. C. Houba (Ulg) & Dr. M. Mergeay (SCK•CEN), > 30 years of research in SCK, studied > 30 labs all over the world



e.g. Toxicity and Reduction testing



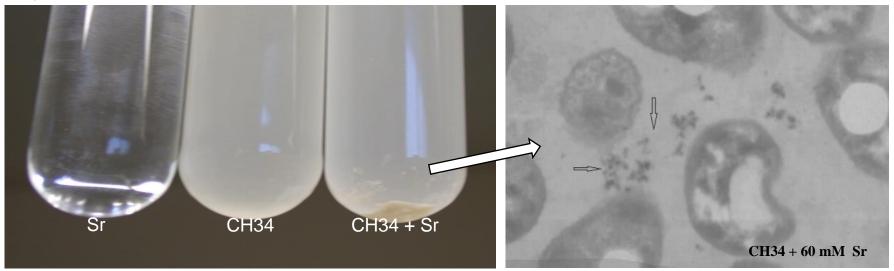
e.g. Biogenic metal nanoparticles

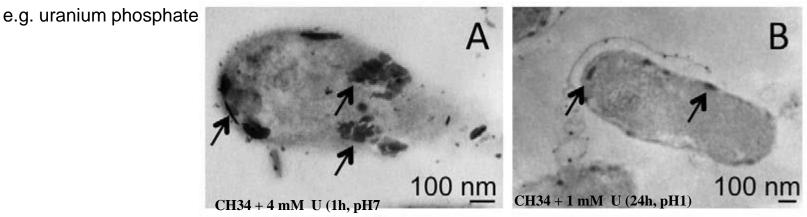


 \rightarrow silver, paladium, gold,

e.g. Biogenic metal carbonates and phosphates

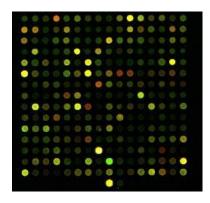
e.g. strontium carbonate

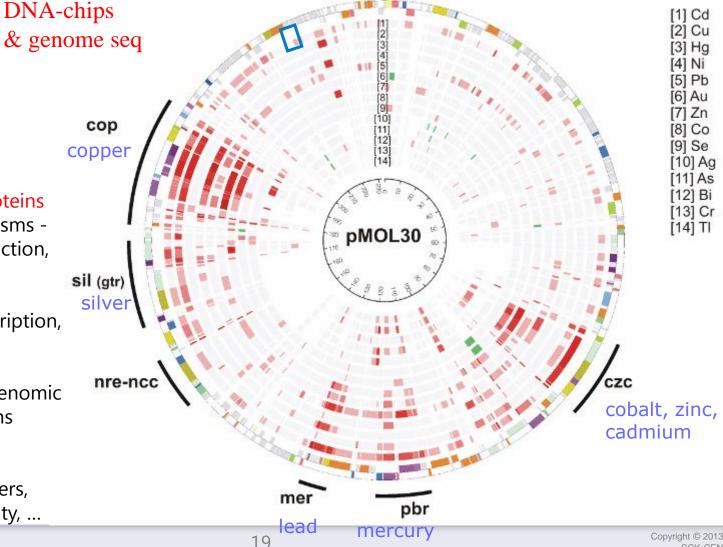




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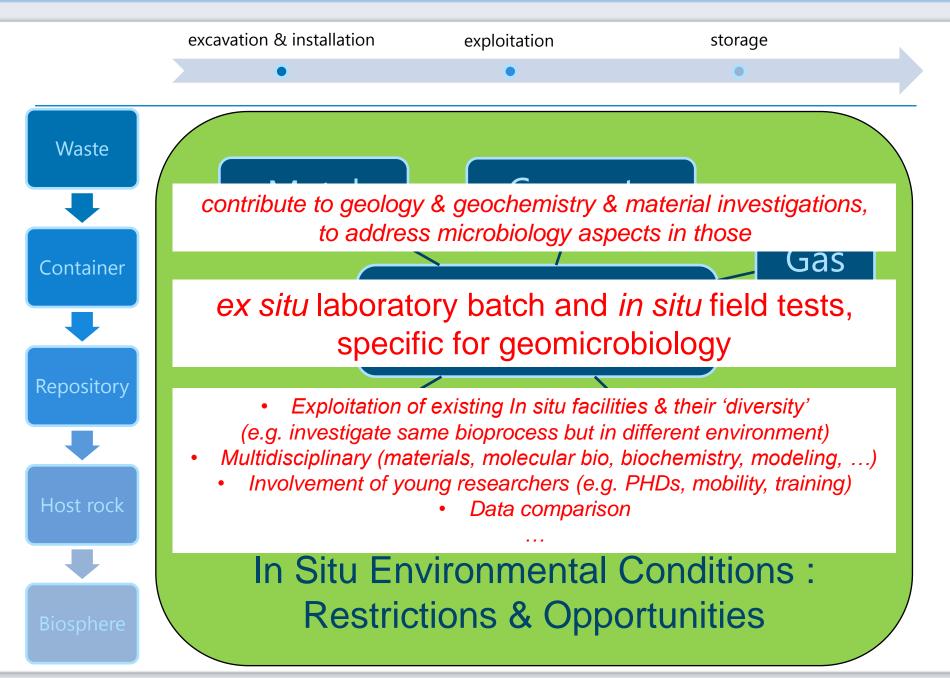
e.g. Characterising genes and proteins in bacteria involved in the metal processing





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- Novel genes & proteins involved : mechanisms efflux pumps, reduction, metallophores ...
- **Regulation**: transcription, regulatory RNA, ...
- Mobility : MGE genomic islands, transposons plasmids, IS, ...
- **Bioreporters** : makers, specificity, sensitivity, ...



We are ready to jump on the new scientific challenges