Microbial influence on the immobilization of radionuclides in crystalline rock environments

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Microbial influence on radionuclide immobilization

- In nonporous crystalline rock, the dominant transport medium for RN is groundwater flowing through subsurface fractures.
- Fracture surfaces support biological growth.
 - Microbial diversity of biofilms depends on microbial consortia and chemical composition of the fracture water.
 - The microbial consortia can significantly affect subsurface biogeochemical interactions, leading to adsorption of RN.

ONKALO: 20 % U uptake by microorganisms, forming hardly soluble minerals Äspö: 85 % U and 95 % Np uptake by bacteriogenic iron oxide precipitates (BIOS)

Subsurface biofilms have a significant effect on the adsorption capacity of host rock formations by forming a barrier between the rock surface and the groundwater.

Biofilms growing on fracture surfaces decrease the rock adsorption capacity for migrating RN.

Biofilm coverage can not be ignored



Example 1:

Future permanent nuclear-waste repository ONKALO, Finland





Intracellular formation of U-phophate mineral



Aqueous uranyl carbonate species in the groundwater



Formation of aqueous uranyl carbonate species in the U contaminated groundwater, most likely $Ca_2UO_2(CO_3)_3$ and $MgUO_2(CO)_3^{2+}$.



Example 2: Hard Rock Laboratory Äspö, Sweden





U and Np uptake in BIOS biofilms

	BIOS biofilm dry mass
	after the experiment with U(VI)
	[µg/g]
Na	1,17E+04
Mg	4,10E+04
AI	9,45E+01
Si	1,01E+03
Р	4,01E+03
к	7,71E+02
Ca	2,06E+04
Fe	3,76E+05
Co	2,29E-01
Ni	3,54E+00
Cu	2,34E+00
As	<5
Br	1,55E+02
Sr	6,19E+02
Sn	4,07E-01
Ва	5,74E+02
Pb	4,24E-01
U	5,00E+02

Precipitation of ferrihydrite onto the extracellular stalks of Gallionella ferruginea depends on:

- total supply of dissolved ferric iron
- availability of reactive sorption sites (i.e. carboxyl or phosphoryl groups) in individual cells



- Substantial uptake of U and Np in BIOS biofilms depends on:
 - high amount of precipitated ferrihydrite (70 % in BIOS biofilm)
 - neutral pH

Uptake of Uranium (85%) and Neptunium (95%) by BIOS biofilms in neutral pH range.



Mobile calcium uranyl carbonate species in the groundwater



* The database used was the thermo.dat accompanying the code, supplemented by the most recent NEA database for aqueous uranium species and mineral uranium phases (Guillaumont et al. 2003 *Chemical Thermodynamics* 5) and Bernhard et al. (2001) *Radiochem. Acta* 89, 511-518.



Expectation on the outcome of WG 5

- Exceptance of the community, that the influence of microbial processes cannot be ignored and have to be implemented in the safety assessment.
- Cooperation with members of WG 5 to expand the knowledge on microbial influence on RN migration behaviour.
 - Meetings in WGs for exchange of knowledge
 - Establishing on-site experiments (e.g. in crystalline rock environments HRL Äspö) under aerobic and anaerobic conditions using long-lived RN
- Bringing forward new research projects



Relevance of microbial processes in a repository environment?

Kinetics of biofilm growth?

Modelling:

- Trace the effect of microbial metabolism and growth in a geochemical system, using a generalized kinetic rate law.
- Estimating biofilm growth for process description, including different scales and different types of radionuclides.



Geochemist's Workbench example calculation



Quantification of microbial processes



Assessment of microbial mediated retention of radionuclides



Remaining issues

What will be the future microbial diversity in a repository in the case of:

- changing geochemical conditions (E_h , pH, T, p)
- opened and closed deposit
- aerobic and anaerobic conditions?
- Which will be the sources for electron donors and electron acceptors for microbial activities?
 - in the far-field
 - in the near-field
- How do microbes influence the corrosion of container materials?
- What is the influence of microbial metabolic products?
- Which parameter are needed for modeling the kinetic of microbial growth?



How can HZDR contribute to a competent safety case?

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- Molecular analyses of the microbial diversity (PCR, cloning, sequencing, pyro-sequencing, FISH techniques)
- Identification of the metabolic activity (e.g. CARD-FISH and RNA analyses)
- Culturing of representative microbial model systems in the laboratory
- Confocal laser scanning microscopy combined with Laser induced fluorescence spectroscopy (CLSM / LIFS)
 RN interactions in situ, speciation in micrometer scale
- Time-resolved Laser-induced Fluorescence Spectroscopy (TRLFS) RN speciation









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How can HZDR contribute to a competent safety case?

- Electron microscopy methods (SEM)/EDS, TEM/EELS)
 ultra structure and elemental composition
- Microelectrode studies for redox determination
 gradients of physico-chemical parameters
- Extended X-ray absorption fine structure spectroscopy (EXAFS)
 structural information on RN













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Concluding remarks for the round table

- For a comprehensive long-term safety analyses it is essential to include microbiological aspects.
- It is necessary to develop a framework to address the microbiological influence relevant for repositories.
- Collaborations and exchange of knowledge as well as establishing of onsite experiments are needed.

Open questions related to repositories and microorganisms:

- How do MO influence the transport of radionuclides?
- How can we define the kinetic of microbial growth?
- How do MO influence the oxidation/corrosion of container materials?
- How do MO influence the stability of reactive barrier material, e.g. bentonite?



Thanks to you for listening patiently!





Henry Moll's presentation this afternoon according to microbes in clay and salt host rocks.

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